

# A molecular framework for the phylogeny of the Pseudocerotidae (Platyhelminthes, Polycladida)

M. K. Litvaitis<sup>1</sup> & L. J. Newman<sup>2</sup>

 <sup>1</sup>Department of Zoology and Center for Marine Biology, Rudman Hall, University of New Hampshire, Durham, NH-03824, U.S.A.
Tel: +603-862-2102. Fax: +603-862-3784. E-mail: MKL1@cisunix.unh.edu
<sup>2</sup>School of Resource Science and Management, Southern Cross University, Lismore, NSW, Australia

Received 6 November 2000; in revised form 6 November 2000; accepted 21 November 2000

Key words: flatworms, phylogeny, 28S rDNA, D3 expansion segment

#### Abstract

Systematic relationships within the cotylean family Pseudocerotidae were examined using nucleotide sequences of the D3 expansion segment of the 28S rDNA gene. A previously suggested separation of *Pseudoceros and Pseudobiceros* based on the number of male reproductive systems was confirmed. Regardless of the algorithm employed, *Pseudoceros* always formed a monophyletic clade. *Pseudobiceros* appeared to be paraphyletic; however, a constrained maximum parsimony tree was not significantly longer (2 steps,  $\alpha = 0.05$ ). Additionally, the genera *Maiazoon, Phrikoceros* and *Tytthosoceros* were validated as taxonomic entities, and their relationships to other genera within the family were determined. Molecular data also supported species separations based on colour patterns. An intraspecific genetic distance of 1.14% was found for *Pseudoceros bifurcus*, whereas the intrageneric distance was 3.58%. Genetic distances among genera varied, with the closest distance being 2.048% between *Pseudobiceros* and *Tytthosoceros*.

## Introduction

Based on the character 'presence/absence of a cotyl or sucker', Lang (1888) divided the order Polycladida into two suborders, the Cotylea and the Acotylea. The systematics of polyclads was reviewed simultaneously by Faubel (1983, 1984) and Prudhoe (1985). Unfortunately, these reviews resulted in two nonconcordant systematic schemes. Prudhoe (1985, 1989) following Hyman (1955a,b,c; 1959a,b), maintained that many species, especially within the colourful Pseudocerotidae, could be diagnosed solely on the basis of colour patterns. Faubel (1984) disagreed and stated that, as is common for other turbellarian flatworms, species should be separated on details of their reproductive structures. Inherent problems surfaced because both reviews relied strongly on the past literature and type material was not located or re-examined. To date, no one system is reliably used and the systematics of these turbellarians is accordingly, in a state of confusion (Newman & Cannon, 1994a).

The cotyleans, with 10–16 families (depending on author), are prominent and colourful members of reef communities (Cannon, 1986). The family Pseudo-cerotidae is the largest and most diverse within the Cotylea. To date, there are approximately 13 genera in the family with an estimated 500 or more species worldwide (Faubel, 1984; Newman & Cannon, 1994a,b, 1996a,b; 1997, 1998).Until Faubel's (1984) revision, *Pseudoceros* with about 75% of the named species in the family, represented the largest genus. Based on the character 'double male reproductive system', Faubel (1984) separated the genus *Pseudobiceros* from *Pseudoceros*.

However, male and female reproductive systems show a surprising homogeneity (Newman & Cannon, 1994a, 1995, 1998a,b), thus, separating species within each genus continued to be difficult. Both genera reproduce via random hypodermic insemination through the body wall (Newman & Cannon, 1994a; Michiels & Newman, 1998). Such a reproductive mode provides a convincing explanation for the relative reproductive homogeneity in these flatworms. Certainly, a reproductive behaviour involving random deposition of sperm through the body wall is unlikely to generate elaborate morphological copulatory isolating mechanisms (Newman & Cannon, 1994a).

Newman & Cannon (1994a, 1995) were able to differentiate 64 new pseudocerotid species based on colour patterns. Unlike earlier descriptions, these authors examined live material and their studies were greatly aided by a new fixation technique that allowed for the preservation of pattern colour (Newman & Cannon, 1995). Furthermore, Newman & Cannon (1994a) showed that individuals of like patterns copulated simultaneously, thus mitigating against any argument of colour pattern polymorphisms within species. Additional support for species distinctiveness is seen in differences between size at maturity and in habitat (Newman & Cannon, 1994a) which indicates that species separated on colour pattern are reliable biological entities. Thus, while genera can be separated on the basis of the male reproductive system, species distinctions within a genus can rely on colour patterns.

Based on the shape of pseudotentacles, pharynx and reproductive anatomy, Newman & Cannon (1996a,b) erected four new pseudocerotid genera, Bulaceros, Maiazoon, Phrikoceros and Tytthosoceros (Table 1). As with Pseudoceros, the genera Phrikoceros and Tytthosoceros possess one male pore but they can be separated from Pseudoceros on morphological differences such as the shape of their pharynx and pseudotentacles, and the arrangement of their eyes. Only Maiazoon is similar to Pseudobiceros in possessing two male reproductive systems, simple folds of the pharynx, and deep marginal ruffling. However, it is separated from that genus by having three to five female antra. Phrikoceros, on the other hand, shares a single male reproductive system with Pseudoceros and is distinguished from that genus by folded pseudotentacles, deep marginal ruffles, and clustered dorsal and ventral pseudotentacular eyes (Newman & Cannon, 1996). To date, no independent validation of these genera has been made.

Molecular phylogenetic studies have become the standard for providing an independent test of existing morphology-based phylogenies (Halanych et al., 1995; Winnepenninckx et al., 1995; Giribet et al., 1996; Aguinaldo et al., 1997; Carranza et al., 1997; Balavoine, 1998; Adoutte et al., 2000 and references therein). In a first attempt to evaluate the usefulness of nucleotide sequence data in resolving pseudocerotid relationships, Goggin & Newman (1996) sequenced about 400 base pairs of the internal transcribed spacer (ITS1) region of the rRNA in three species of *Pseudoceros*. These authors found sufficient variation to unequivocally discriminate among the three species.

We therefore wanted to evaluate the usefulness of the D3 expansion segment of the 28S rDNA gene for the phylogenetic resolution of pseudocerotid flatworms. This segment has previously been shown to be of phylogenetic value in resolving relationships at various taxonomic levels, ranging from species to class (Litvaitis et al., 1994, 1996, 2000; Nunn et al., 1996). Our specific objectives were (1) to determine if separating *Pseudoceros* and *Pseudobiceros* based on the number of male reproductive systems is confirmed by molecular data, (2) to determine if species separated according to the colour pattern grouping system established by Newman & Cannon (1994a) are valid entities, and (3) to provide a first test of the validity of the genera *Maiazoon, Phrikoceros* and *Tytthosoceros*.

## Materials and methods

Cotylean flatworms were hand collected from the Great Barrier Reef (GBR), Australia; Papua New Guinea and Dominica, West Indies (Table 2). DNA was extracted according to Litvaitis et al. (1994, 1996), and amplified using primers designed to conserved regions around the D3 expansion segment of the gene coding for 28S rDNA (for primer sequences, see Litvaitis et al., 1994). Amplified products were gel purified, and 4–5  $\mu$ l of each sample were used in a cycle-sequencing reaction (protocol according to ABI Inc.). Fragments were separated on a 6% denaturing polyacrylamide gel, and sequences were determined using an automated sequencer (ABI 377). Initial editing of sequences was done via the SeqEd program (version 1.0.1; ABI Inc.). Although the products were only about 350 base pairs long, both strands were sequenced to assure accuracy.

Sequences were aligned by the CLUSTAL method, using MegAlign (DNA\*) with further improvements of the alignment by eye. An initial neighbor joining tree (NJ) was produced (PAUP\*; Swofford, 1999) using two specimens of acotyleans and the macrostomid *Microstomum papillosum* as an outgroup. Macrostomids have been shown to represent the immediate sister group of polyclads (Carranza et al., 1997; Litvaitis & Rohde, 1999). To correct for multiple substitutions, data were log/Det transformed for NJ-

a			<b>D</b>		<b>D1</b> 11		
Character	Acanthozoon	Pseudoceros	Pseudobiceros	Maiazoon	Phrikoceros	Thysanozoon	Tytthosoceros
Body shape	raised medially	flat	raised medially	raised medially	raised medially	raised medially	raised medially
Dorsal surface	papillate	smooth	smooth	smooth	smooth	papillate	smooth
Pseudotentacle shape	<sup>a</sup> ear-like	simple	ear-like or square	square	square	<sup>a</sup> ear-like	ear-like
Pharyngeal folds	<sup>a</sup> complex	complex	simple	simple	simple	<sup>a</sup> simple	simple
Cerebral eye clusters	<sup><i>a</i></sup> 4 clusters	anterior lines	4 clusters	4 clusters	4 clusters	<sup><i>a</i></sup> 4 clusters	scattered
Number of female pores	1	1	1	3–5	1	1	1
Number of male pores	1	1	2	2	1	2	1

Table 1. Summary of generic diagnostic characters for selected Pseudocerotidae (Faubel, 1984; Prudhoe, 1985; Cannon, 1986; Newman and Cannon, 1994a, 1996a). Note only genera used in this study are listed

<sup>a</sup>Newman, unpublished data.

trees. Reliability of internal nodes was ascertained by 2000 bootstrap replications. A maximum parsimony (MP) analysis using heuristic search was conducted with random sequence addition and tree bisectionreconnection branch swapping (PAUP\*; Swofford, 1999). An alternative MP hypothesis was evaluated where Pseudobiceros was constrained into a monophyletic clade. Using the non-parametric ranked sign test of Templeton (Larson, 1994) at  $\alpha = 0.05$ , it was shown that the constrained tree was not significantly longer (2 additional steps). Alternative longer trees that provided a better concordance with morphological characters are favored as long as they are not statistically different from the MP tree (Litvaitis & Rohde, 1999, Litvaitis et al., 2000). We therefore used the constrained MP tree in our analysis.

# **Results and discussion**

Regardless of the algorithm employed, the genus *Pseudoceros* always formed a distinct clade (Figs 1 and 2). The coherence of the genus is further supported by morphological characters associated with the eyes, pseudotentacles, pharynx and by details of the reproductive system (Newman & Cannon, 1994a). Within the genus, three specimens of *Pseudoceros bi-furcus* formed a clade whose sister group included two



*Figure 1.* Neighbor-joining tree of 25 polyclad specimens, using partial sequences of the 28S rDNA gene (D3 expansion segment). Numbers at nodes are percentages of 2000 bootstrap replications; only values above 50% are reported.

Table 2. Species and collection localities

Taxon	Collection locality		
Pseudoceros			
bicolor	Dominica, West Indies		
bifurcus A, B	North Heron Island, GBR, Queensland,		
	Australia		
bifurcus D	Blue Pools, North Heron Island, GBR,		
	Queensland, Australia		
irretitus	North Heron Island, GBR, Queensland,		
	Australia		
paralaticlavus	North Heron Island GBR Queensland		
Puluiuleiu (ub	Australia		
rubronanus	North Heron Island GBR Queensland		
rubronanus	Australia		
aannhininya	Ausualia North Horon Joland, CBB, Queensland		
sappiirmus	North Heron Island, OBR, Queensland,		
11 / A 1D	Austrana		
unident. sp. A and B	Point Cartwright, Mooloolabe, SE		
	Queensland, Australia		
Pseudobiceros			
bedfordi	Blue Pools North Heron Island GRP		
bearbrai	Queensland Australia		
arotue	North Heron Island GBR Oueensland		
Siddas	Australia		
hanaaakanua	Haron Island CPP Quaanaland		
hancockanus	Australia		
	Ausuana Madana Dama Nam Cainaa		
uniarborensis A	Madang, Papua New Guinea		
uniarborensis B	North Heron Island, GBR,		
	Queensiand, Austrana		
Acanthozoon			
unidentified sp	Madang Papua New Guinea		
undentified sp.	Madang, Fupua Now Sumea		
Maiazoon			
orsaki	Madang, Papua New Guinea		
Phrikoceros			
galacticus	Heron Island, GBR, Queensland,		
	Australia		
Inysanozoon			
unidentified sp. A and B	Madang, Papua New Guinea		
T-441			
1 yttnosoceros	fillerer blad CDD O		
nocturnus	off Heron Island, GBR, Queensland,		
	Australia		
Doricolis			
unidentified on A	Madang Danua Naw Crimes		
undenuned sp. A	Ligand Joland, CDD, Output		
undenuned sp. B	Australia		
	Ausudilä		



*Figure 2.* Consensus MP tree of 25 polyclad specimens, using partial sequences of the 28S rDNA gene (D3 expansion segment). Numbers at nodes are percentages of 2000 bootstrap replications; only values above 50% are reported. Note, *Pseudobiceros* was constrained as a monophyletic group (\*). (Tree length 184; CI = 0.723; RI = 0.702; RC = 0.507).

unidentified species of *Pseudoceros* (A and B). The average genetic distance among the three specimens of *P. bifurcus* was 1.14%. Using this value as a measure of intraspecific variation, we compared genetic distances among different *Pseudoceros* species that had been separated based on colour patterns. A genetic distance value of 0.609% between *Pseudoceros* sp. A and *Pseudoceros* sp. B led us to conclude that the two specimens were very likely the same species. Both specimens were cream-coloured. Although one was surrounded by a continuous purple margin, whereas the margin of the second specimen was discontinuous and slightly more blue in colour, these differences fall well within the range of variation observed for other pseudocerotid colour patterns.

A comparison of genetic distances of the two unidentified *Pseudoceros* species with *P. bifurcus* D (1.651% and 1.636%, respectively) showed that the two may be closely related to *P. bifurcus*, a conclusion that was further supported by 100% bootstrap replications (Fig. 1). The average genetic distance within the genus *Pseudoceros* was 3.58%, indicating that species separations based on colour patterns are valid

Acanthozoon	Maiazoon	Phrikoceros	Pseudobiceros	Pseudoceros	Thysanozoon	Tytthosoceros	
Acanthozoon							
Maiazoon	0.02052	_					
Phrikoceros	0.03483	0.03044	_				
Pseudobiceros	0.02630	0.02048	0.02354	—			
Pseudoceros	0.05162	0.04632	0.04707	0.04101	—		
Thysanozoon	0.04997	0.05856	0.05462	0.04281	0.06951	_	
Tytthosoceros	0.05584	0.06932	0.06106	0.06369	0.08345	0.08220	_

Table 3. LogDet/paralinear distance matrix of pseudocerotid genera. Values for Thysanozoon, Pseudoceros and Pseudobiceros, represent intrageneric averages

and allow for a reliable discrimination of individual species.

The genus Pseudobiceros proved to be a more heterogeneous group (Fig. 1). Unconstrained analyses resulted in a paraphyletic genus with most of the examined species grouping with members of Thysanozoon and two specimens of Pseudobiceros uniarborensis clustering with Phrikoceros galacticus. A close relationship between Pseudobiceros and Thysanozoon is supported though by both genera possessing two male pores and the same pharynx structure. When Pseudobiceros was constrained (MP analysis) into a monophyletic clade, the resulting tree was not significantly longer (2 additional steps,  $\alpha = 0.05$ ) as determined by the non-parametric ranked sign test of Templeton (Larson, 1994). More importantly though, constraining a monophyletic Pseudobiceros did not influence the topology of relationships within Pseudoceros (Fig. 2). As a consequence, Pseudobiceros should be considered a valid genus, distinct from Pseudoceros, Thysanozoon or Phrikoceros. Additionally, we recommend that alternative trees that are supported by morphological characters be favoured, as long as they are not statistically different from the MP trees. The genetic distance between the two specimens of P. uniarborensis was 1.25%, a value that was comparable to the intraspecific distance of Pseudoceros bifurcus.

An examination of intergeneric distances (Table 3), as expected, resulted in larger values than val-

ues within a genus. For example, genetic distances between *Acanthozoon* and *Tytthosoceros, Maiazoon*, and the average of the two *Thysanozoon* specimens was 5.584%, 2.052% and 4.997%%, respectively (Table 3). These values were comparable to values common for other flatworms (Litvaitis et al., 1994; Litvaitis & Rohde, 1999). Not surprisingly, the closest intergeneric distance was found between *Maiazoon* and *Pseudobiceros* (2.048%). The two genera are both characterized by two male reproductive systems, the same type of simple pharyngeal folds, and a similar marginal ruffling (Table 1). However, characters of the female reproductive system clearly separate them into two genera (Newman & Cannon, 1996).

Comparisons of genetic distances for *Phrikoceros* and *Maiazoon* with the remaining genera examined in this study, revealed their distinctness (Table 3). *Phrikoceros*, which shares characters with Pseudoceros and Pseudobiceros, was more closely related to *Pseudobiceros* (2.354%); however, this value was still higher than the intrageneric values determined for *Pseudobiceros*.

Using a morphology independent data set, the present study confirmed and validated the genera *Pseudoceros* and *Pseudobiceros* as taxonomic entities. Additionally, we were able to confirm the use of colour patterns to distinguish species. Species identifications based on nucleotide sequence although unequivocal, result in the destruction of the specimen. Thus, the colour scheme of Newman & Cannon (1994a, 1995)

which preserves specimens intact, is a significant and valid contribution to polyclad systematics. Finally, the present study also supported the establishment of *Phrikoceros, Maiazoon* and *Tytthosoceros* as separate genera (Newman & Cannon, 1996) and their relationships to other genera within the family Pseudocerotidae.

### Acknowledgements

We thank the many people who helped collect polyclads, especially Dr A. Flowers and Mr W. Ellis. Special thanks is given to the Directors and staff at Heron and Lizard Island Research Stations, the Department of Invertebrates of the National Museum of Natural History, Smithsonian Institution, and the School of Resource Science and Management, Southern Cross University, NSW. The Australian Biological Resource Study, Canberra, Australia, the National Museum of Natural History, Washington, DC, and the Christensen Research Institute, Madang, Papua New Guinea generously provided financial support to L. N. This study was funded by a grant from the Hubbard Marine Research Initiation Program at the University of New Hampshire.

#### References

- Adoutte, A., G. Balavoine, N. Lartillot, O. Lespinet, B. Prud'homme & R. De Rosa, 2000. The new animal phylogeny: Reliability and implications. Proc. Nat. Assoc. Sci. 97: 4453–4456.
- Aguinaldo, A. M. A., J. M. Turbeville, L. S. Linford, M. C. Rivera, J. R. Garey, R. A. Raff & J. A. Lake, 1997. Evidence for a clade of nematodes, arthropods and other moulting animals. Nature 387: 489–493.
- Balavoine, G., 1998. Are Platyhelminthes coelomates without a coelom? An argument based on the evolution of *Hox* genes. Am. Zool. 38: 843–858.
- Cannon, L. R. G., 1986. Turbellaria of the World A Guide to Families and Genera. Queensland Museum, Brisbane: 136 pp.
- Carranza, S., J. Baguña & M. Riutort, 1997. Are the Platyhelminthes a monophyletic primitive group? An assessment using 18S rDNA sequences. Mol. Biol. Evol. 14: 485–497.
- Faubel, A., 1983. The Polycladida, Turbellaria. Proposal and establishment of a new system. Part I. The Acotylea. Mitt. Hamb. Zool. Mus. Inst. 80: 17–121.
- Faubel, A., 1984. The Polycladida, Turbellaria. Proposal and establishment of a new system. Part II. The Cotylea. Mitt. Hamb. Zool. Mus. Inst. 80: 189–259.
- Goggin, C. L. & L. J. Newman, 1996. Use of molecular data to discriminate pseudocerotid turbellarians. J. Helminth. 7: 123– 126.
- Halanych, K. M., J. D. Bacheller, A. M. A. Aguinaldo, S. M. Liva, D. M. Hillis & J. A. Lake, 1995. Evidence from 18S ribosomal DNA that the lophophorates are protostome animals. Science 267:1641–1643.

- Hyman, L. H., 1955a. Some polyclads from Polynesia and Micronesia. Proc. U.S. Natl. Mus. 105: 65–82.
- Hyman, L. H., 1955b. Some polyclads from the West Indies and Florida. Proc. U.S. Natl. Mus. 104: 115–150.
- Hyman, L. H., 1955c. A further study of the polyclad flatworms of the West Indian region. Bull. mar. Sci. Gulf Carib. 5: 259–268.
- Hyman, L. H., 1959a. A further study of Micronesian polyclad flatworms. Proc. U.S. Natl. Mus. 108: 543–597.
- Hyman, L. H., 1959b. Some Australian polyclads. Records Australian Museum 25: 1–17.
- Larson, A., 1994. The comparison of morphological and molecular data in phylogenetic systematics. In Schierwater, B., B. Streit, G. B. Wagner & R. DeSalle (eds), Molecular Ecology and Evolution: Approaches and Applications. Birkhäuser Verlag, Basel: 371–390.
- Litvaitis, M. K. & K. Rohde, 1999. A molecular test of platyhelminth phylogeny: inferences from partial 28S rDNA sequences. Invert. Biol. 118: 42–56.
- Litvaitis, M. K., G. Nunn, K. Thomas & T. D. Kocher, 1994. A molecular approach for the identification of meiofaunal turbellarians (Platythelminthes, Turbellaria). Mar. Biol. 120: 437–442.
- Litvaitis, M. K., M. C. Curini-Galletti, P. M. Martens & T. D. Kocher, 1996. A reappraisal of the systematics of the Monocelididae (Platyhelminthes, Proseriata): inferences from molecular data. Mol. Phylog. Evol. 6: 150–156.
- Litvaitis, M. K., J. W. Bates, W. D. Hope & T. Moens, 2000. Inferring a classification of the Adenophorea (Nematoda) from nucleotide sequences of the D3-expansion segment (26/28S rDNA). Can. J. Zool. 78: 911–922.
- Michiels N. & L. J. Newman, 1998. Sex and violence in a hermaphrodite. Nature 391(6668): 647
- Newman, L. J. & L. R. G. Cannon, 1994a. Pseudoceros and Pseudobiceros (Polycladida, Pseudocerotidae) from Eastern Australia and Papua New Guinea. Mem. Qld. Mus. 37: 205–266.
- Newman, L. J. & L. R. G. Cannon, 1994b. Biodiversity of Australian polyclad flatworms. Mem. Qld. Mus. 36: 159–163.
- Newman, L. J & L. R. G. Cannon, 1995. The importance of the fixation of colour, pattern and form in tropical Pseudocerotidae (Platyhelminthes, Polycladida). Hydrobiologia 305: 141–143.
- Newman, L. J. & L. R. G. Cannon, 1996a. New genera of pseudocerotid flatworms (Platyhelminthes, Polycladida) from Australian and Papua New Guinean coral reefs. J. Nat. Hist. 30: 1425–1441.
- Newman, L. J. & L. R. G. Cannon, 1996b. *Bulaceros*, new genus and *Tytthosoceros*, new genus (Platyhelminthes, Polycladida, Pseudocerotidae) from the Great Barrier Reef, Australia and Papua New Guinea. Raffles Bull. Zool. 44: 479–492.
- Newman, L. J. & L. R. G. Cannon, 1997. Nine new *Pseudobiceros* (Platyhelminthes, Polycladida, Pseudocerotidae) from the Indo-Pacific. Raffles Bull. Zool. 45: 341–368.
- Newman, L. J. & L. R. G. Cannon, 1998. New *Pseudoceros* (Platyhelminthes, Polycladida, Pseudocerotidae) from the Indo-Pacific. Raffles Bull. Zool. 46: 293–323.
- Nunn, G. B., B. F. Theisen, T. B. Christensen & P. Arcanter, 1996. Simplicity-correlated size growth of the nuclear 28S ribosomal RNA D3 expansion segment in the crustacean order Isopoda. J. Mol. Evol. 42: 221–223.
- Prudhoe, S., 1985. A Monograph on Polyclad Turbellaria. British Museum (Natural History), Oxford Univ. Press, Oxford: 259 pp.
- Prudhoe, S., 1989. Polyclad turbellarians recorded from African waters. Bull Brit. Mus. Nat. Hist. (Zool) 55: 47–96.
- Swofford, D., 1999. PAUP\*: Phylogenetic Analysis Using Parsimony (and other methods), vers. 4.0, Sinauer Associates, Sunderland, MA.